

# Interbacterial competition assay

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Antibacterial T6SS effectors with a VRR-Nuc domain are structure-specific nucleases

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## Detailed protocol

### Interbacterial competition assay

#### Day 1

Bacterial strains of *Salmonella bongori* (See Supplementary file 7), and *Escherichia coli* K-12 W3110 pEXT22 Km<sup>R</sup> were streaked in Lysogeny Broth- agar plates (10 g/L tryptone, 10 g/L NaCl, 5 g/L yeast extract, 1.5% agar) – 90mm plates with 25 mL of medium with additional antibiotics as needed. All strains were grown for 12-16 h (overnight, ON) under 200 rpm at 37 °C.

#### Day 2

Single colonies of bacterial strains were inoculated in 3 mL of LB with appropriate antibiotics in 15 mL tubes and grown ON at 37 °C under 200 rpm.

#### Day 3

ON cultures of *S. bongori* (WT,  $\Delta tssB$ ,  $\Delta tssB$  pFPV25.1 *tssB* Amp<sup>R</sup>,  $\Delta vgrG1$ ,  $\Delta vgrG2$ , or  $\Delta vgrG3$ ) as attackers and *E. coli* K-12 W3110 pEXT22 Km<sup>R</sup> or *S. bongori* ( $\Delta tseV2/tseV2.1/tseV2.2$  Km<sup>R</sup>,  $\Delta tseV2/tseV2.1/tseV2.2$  Km<sup>R</sup> pFPV25.1 *tseV2.1*,  $\Delta tseV3/tseV3$  Km<sup>R</sup> or  $\Delta tseV3/tseV3$  Km<sup>R</sup> pFPV25.1 *tseV3*) as prey were subculture in 5 mL of LB (1:30 dilution) in a 50 mL tube. Cultures were grown at 37 °C under 200 rpm until reaching OD<sub>600 nm</sub> 1.6, then adjusted to OD<sub>600 nm</sub> 0.4 and mixed with vortex in a 10:1 ratio (attacker:prey) in a 1.5 mL tube (200  $\mu$ L:20  $\mu$ L).

**Input preparation** –5  $\mu$ L of the mixture were transfer to a 1.5 mL tube with 995  $\mu$ L of LB medium and homogenized by vortex. Serial dilutions were prepared by transferring 100  $\mu$ L to 900  $\mu$ L of LB up to 10<sup>3</sup> and plated on LB agar plates with antibiotics for prey selection. Plates for input were grown ON at 37 °C. CFU (colony-forming units) were recorded.

**Output preparation** – 1x1 cm nitrocellulose membranes (pore 0.22  $\mu$ m) were place on a sterile 90 mm petri dish and 5  $\mu$ L of the mixture were spotted onto the membrane. After the mixture was absorbed by the membranes, these were immediately transferred and positioned face-up in a 90 mm plates with 40 mL of LB-agar - carefully to avoid bubble formation between medium and membrane. The membranes for output were incubated at 37°C with the membrane facing up. After the estimated time of competition, the membranes were transfer from the plates to a 1.5 mL tube with 1 mL of LB medium and homogenized by vortex. Serial dilutions were prepared up to 10<sup>5</sup> and plated on 25 mL LB agar plates with antibiotics for prey selection. Plates for output were grown overnight at 37 °C. CFU counts were recorded.

The prey recovery rate was calculated by dividing the CFUs counts of the output by the CFU of the input. The rates were normalize by the values recovered for *S. bongori* WT as control. One-way analysis of variance (ANOVA) followed by Dunnett's multiple comparison test. \*\*p < 0.01, and \*\*\*p < 0.001.

**How to cite:** (Readers should cite both the Bio-protocol preprint and the original research article where this protocol was used)

1. Bayer-Santos, E. and Sanchez-Limache, D. E.(2023). Interbacterial competition assay. Bio-protocol Preprint. [bio-protocol.org/prep2287](https://bio-protocol.org/prep2287).
2. Hespanhol, J. T., Sanchez-Limache, D. E., Nicastro, G. G., Mead, L., Llontop, E. E., Chagas-Santos, G., Farah, C. S., de Souza, R. F., Galhardo, R. D. S., Lovering, A. L. and Bayer-Santos, E.(2022). Antibacterial T6SS effectors with a VRR-Nuc domain are structure-specific nucleases. eLIFE. DOI: [10.7554/eLife.82437](https://doi.org/10.7554/eLife.82437)

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